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Calorimetric & Respirometric Monitoring of Metabolism: Some Examples

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Instrument:

2277 Thermal Activity Monitor

Field of Application:

Biology

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Introduction

Simultaneous measurements of the rates of heat production and oxygen utilisation by living tissues provides a direct quantitative means of detecting subtle changes in metabolic state in the face of altered physiological and environmental conditions. The ratio of calorimetrically measured heat flux to the respirometrically measured oxygen flux is often called the "calorimetric-respirometric ratio" or CR ratio. This can be used to partition total metabolic energy flux into its aerobic and anaerobic components by comparison of the experimental CR ratio with the theoretical "oxycaloric equivalent" for fully aerobic respiration. Such information is most important when evaluating the physiological effect of environmental stress.

Industrialisation of many processes has led to an increasing amount of synthetic and natural waste products expelled into the environment. The consequences of this can often be fatal as seen by the many

examples of polluted rivers, waste land and the atmosphere. This large amount of waste can be seen to be harmful to the ecosystem and has led to an imbalance of the natural flora and fauna. There is a need, therefore to study the effects of pollutants and to evaluate quantitatively their toxicological effects. The combined use of microcalorimetry and respirometry provides a useful means of monitoring such effects.

Examples

Dormancy and hibernation are common among many organisms during their lifecycles. These metabolic and physiological changes are beginning to be better understood from a biochemical point of view. Using brine shrimp (*Artemia*) embryos, the role of intracellular pH in metabolic switching was investigated during anaerobic dormancy (Ref 1). *Artemia* embryos placed in a ThermoMetric perfusion vessel coupled to the Cyclobios Twin-Flow respirometer

exhibited an increase in heat flux as a function of embryonic development. They reached steady state after 7h under aerobic conditions (Fig 1). At steady state the CR ratio (calorimetric heat change per respirometric oxygen uptake) was -495 kJ/mol O_2 for fully aerobic carbohydrate metabolism which is close to the oxycaloric equivalent of -478 kJ/mol for carbohydrate catabolism. Exposure of *Artemia* to CO_2 under aerobic conditions resulted in a cessation of development and a decrease in heat flux to 9% of control levels after 8h. Oxygen flux decreased simultaneously, that is the CR ratio remained approximately equal to the oxycaloric equivalent, indicating fully aerobic metabolism. Removal of CO_2 resulted in a rapid increase in heat flux to control levels.

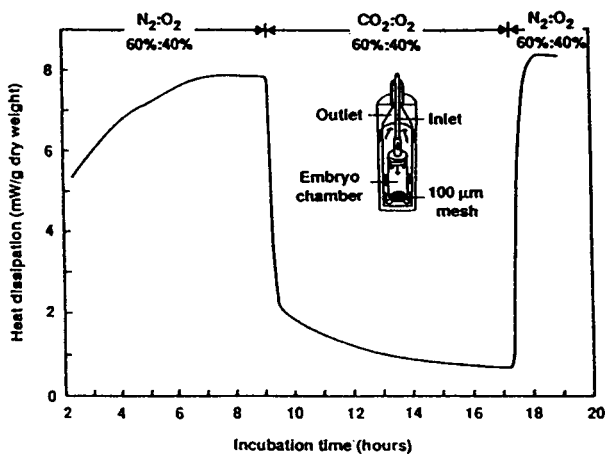


Fig 1. Heat dissipation by *Artemia* embryos under varying aerobic/acidosis conditions (Ref 1).

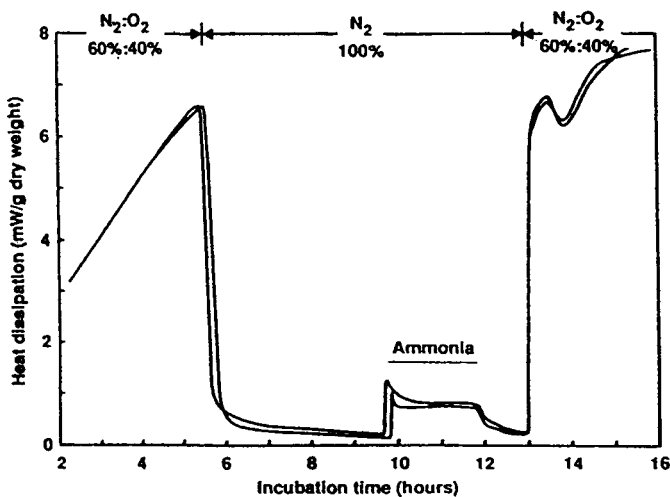


Fig 2. Heat dissipation by *Artemia* embryos under aerobic and anaerobic conditions. Dormancy was interrupted by addition of ammonia which was later removed (Ref 2).

In other experiments, oxygen was replaced by 100% nitrogen during the aerobic development of *Artemia*. An immediate drop in heat output was observed which declined to $2.4 \pm 0.4\%$ of control values (Fig 2). Thus showing the degree to which oxygen is required for metabolic activity.

The internal pH is the primary regulatory factor which governs the metabolic pathway used by the *Artemia*. Addition of ammonia to *Artemia* under anaerobic dormancy resulted in an increased anoxic heat output as a consequence of the change in biochemical pathway utilised. However, since oxygen was not present for mitochondrial metabolism to function normally, the heat flux was not as high as control values. Nevertheless, a five fold increase was observed compared to the anaerobic dormant condition. Reinstatement of the oxygen presence caused a rapid bi-phasic increase in heat output to normal pre-anoxic levels.

Many other aquatic organisms such as the common mussel (*Mytilus edulis*) have also been examined for simultaneous oxygen and heat flux measurements (Ref 2). The gills of *Mytilus* were carefully excised and placed in the TAM perfusion vessel under aerobic, hypoxic and anoxic conditions (Ref 3). Cilia on the gills propel water across the gills to provide oxygen and nutrition. The rate of ciliary activity is principally governed by the balance of the rate of formation and utilisation of ATP. Hence, factors that influence the rate of formation of ATP will invariably effect the rate of ciliary activity.

Results from stimulated ciliary activity simultaneously monitored in the TAM connected to the Cyclobios Twin-Flow respirometer (Fig 3) suggest that the rate of utilisation of oxygen and heat output are directly related to the rate of ciliary activity and that the energy metabolism of intact gills is mainly aerobic. Therefore, the presence of pollutants and other such chemicals could be added in order to observe the toxicological effects of these agents.

A number of mammalian cells have also been examined for information on the CR ratio (Ref 4). It has been suggested that a number of cell types always retain a degree of anaerobic metabolism in addition to the normal aerobic contribution. Simultaneous measurement of the oxygen uptake and heat flux has revealed the extent to which this occurs in living cells.

Results from a number of different mammalian cell types (Fig 4) have shown quantitative differences in anaerobic metabolism. This suggests that different

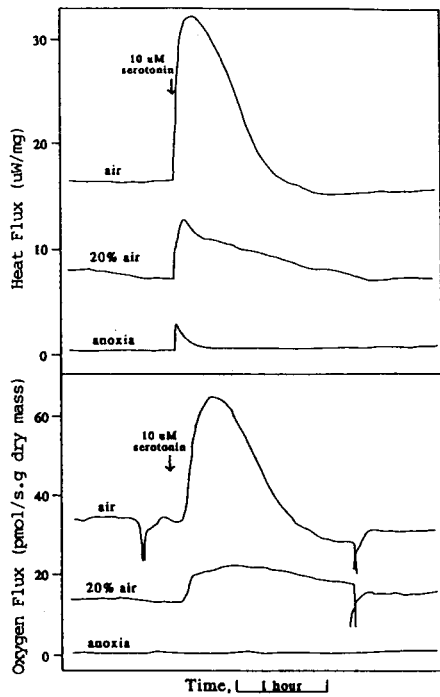


Fig 3. Heat and oxygen flux of *Mytilus edulis* gills under different conditions of air saturation.

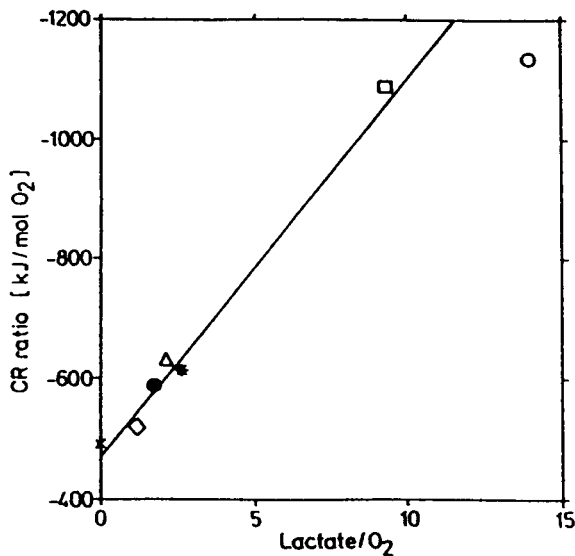


Fig 4. Calorimetric - respirometric (CR) ratio as a function of the molar lactate/ O_2 ratio in cultured or isolated mammalian cells (Ref 4). Hamster brown adipocytes (\times); mouse macrophage hybridoma 2C11-12 (\diamond); human neutrophils activated for oxidative burst (\bullet) and resting (\circ); human T-lymphoma cells CCRF-CEM during growth (\ast) and under non-growing conditions (\square); LS-L929 fibroblasts (\triangle).

metabolic pathways are utilised in the generation of ATP. Hence, it can be further suggested that the CR ratio can be used to differentiate between different cell types under the same physiological conditions.

Conclusion

The above examples have demonstrated the usefulness of both microcalorimetric and oxygen measurements in establishing the effects of different environmental factors upon metabolism. Indeed, the results have shown quantitative differences between different cell types as well as the degree to which the anaerobic contribution exists.

References

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- (3) Doeller, J.E. Kraus, D.W. Gnaiger, E. Shick, J.M. 1990 - Calorespirometry and spectrophotometry of the ciliated gill of the marine mussel *Mytilus edulis*. *Thermochim. Acta* 172:171-178.
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Note

Thermometric AB collaborates with Cyclobios (Austria) in the improvement of Calor-Respirometric systems.